

ISSN : 2309-3234

TMSS MEDICAL COLLEGE JOURNAL

Volume 04, No. 01, January 2015



An Official Publication of TMSS Medical College, Bogra



TMSS Medical College Journal

January 2015

An Official Publication
of
TMSS Medical College, Bogra

TMSS Medical College Journal

Vol.3, No. 1, January 2015

An Official Publication of TMSS Medical College, Bogra, Bangladesh

Editorial Board

Chairperson : Prof. Dr. Most. Rebeca Khatoon
Editor-In-Chief : Prof. Dr. Md. Abdus Shukur
Executive Editor : Prof. Dr. Md. Azfarul Habib
Publication Editor : Dr. Md. Abdul Mazid
Finance Editor : Dr. Md. Matiur Rahman

Published by

Dr. Md. Abdul Mazid
on behalf of the TMSS Medical College

Graphic Designer

Md. Sirajul Islam
TMSS, Bogra

Members

Prof. Dr. M. A. Gafur
Prof. Dr. Md. Aynul Huq
Prof. Dr. Md. Shahjahan Ali Sarkar
Prof. Dr. Nur-A-Farhana Islam
Dr. Md. Azizar Rahman
Dr. Md. Sheik Abdul Muktaf
Dr. Md. Kamrul Islam

Printed by

TMSS Printing Press
Bogra, Bangladesh

Review Committee

Prof. Dr. Raghib Ahsan
Prof. Dr. Md. Anup Rahman Chowdhury
Prof. Dr. Md. Abul Hossain Khan

Advisory Board

Prof. Dr. Hosne-Ara Begum
Prof. Dr. A.K.M Ahsan Habib
Prof. Dr. Syed Mainul Hasan
Prof. Dr. S.A.M. Israrul Bari
Prof. Dr. Md. Rafiqul Islam
Prof. Dr. Latifur Rahman
Prof. Dr. Abdul Gaffar
Prof. Dr. Hafiza Arzuman
Dr. A.K.M. Ahsan Habib
Dr. Md. Mostafa Alam

The TMSS Medical College Journal is a peer reviewed Journal. It is published two times a year, (January and July). It accepts original articles, review articles and case reports. Complimentary copies of the journal are sent to libraries of all medical and other relevant academic institutions in the country and selected institutions abroad.

While every effort is always made by the Editorial Board and the members of the Journal Committee to avoid inaccurate or misleading information appearing in the TMSS Medical College Journal, information within the individual article are the responsibility of its author(s). The TMSS Medical College Journal, its Editorial Board and Journal Committee accept no liability whatsoever for the consequences of any such inaccurate and misleading information, opinion or statement.

Address of Correspondence

Executive Editor, TMSS Medical College Journal
TMSS Medical College, Rangpur Road
Thengamara, Bogra, Bangladesh.
Phone: 051-61830, Fax : 051-61830
e-mail: editortmcjournal@gmail.com
Web: www.tmssmedicalcollege.com

Original Articles**Distribution and Antimicrobial Susceptibility Pattern of Commonest Bacteria Causing Urinary Tract Infection: A Study in A Tertiary Care Hospital of Bangladesh**Khan AH^{1*}, Yasmin T², Rahman A³, Rahman A⁴, Samad A⁵, Akhter S⁶.

1. Dr. Md. Abul Hossain Khan, Professor & Head, Department of Microbiology, TMSS Medical College, Bogra.

2. Dr. Taslima Yasmin, Assistant Professor, Department of Microbiology, North Bengal Medical College, Sirajgonj.

3. Dr. Md. Arifur Rahman, Assistant Professor, Department of Microbiology, TMSS Medical College, Bogra.

4. Dr. Md. Afzalur Rahman, Professor(cc) & Head, Department of Biochemistry, Prime Medical College, Rangpur.

5. Dr. Md. Abdus Samad, Associate Professor & Head, Department of Community Medicine, Prime Medical College, Rangpur.

6. Dr. Shamima Akhter, Assistant Professor, Department of Pathology, TMSS Medical College, Bogra.

* Corresponding Author

Abstract

Background: Urinary tract infection (UTI) is one of the common infections in the community and hospital. Bacterial distribution and susceptibility of UTI changes according to time and place. **Objective:** This study was conducted to determine the bacterial distribution and antimicrobial susceptibility of common uropathogen in a tertiary care hospital of Bangladesh. **Methodology:** Clean catch midstream urine samples were collected from 494 patients irrespective of the age and sex. Isolation of both gram positive and gram negative bacteria by standard culture technique and identification was done on the basis of their cultural and gram staining smear made from culture colonies and biochemical characteristics. Antimicrobial susceptibility was performed in common isolated bacteria by Kirby Bauer's disc diffusion method. **Results:** Among the 494 sample, 212 showed culture positive (42.9%) and Gram negative bacteria (94.3%) were found in high prevalence and *Escherichia coli* (58.96%) was the most prevalent gram negative isolates. Nalidixic acid (83.4%) and ciprofloxacin (88.4%) were found the most resistant and Imipenem and Nitrofurantoin were found the most susceptible drugs against isolated *Escherichia coli* which showed 94.1% and 87.5% susceptibility, respectively. **Conclusion:** *Escherichia coli* is the most common bacteria causing UTI in both sexes and irrespective of age. Imipenem, Nitrofurantoin and Amikacin are the most sensitive, Nalidixic acid, ciprofloxacin, cefuroxime and cephradine are the most resistant antibiotics.

Key words: UTI, Bacterial distribution, antibiotic susceptibility

Introduction

Urinary tract infection (UTI) is the commonest bacterial infectious disease in community practice with a high rate of morbidity and financial cost. It has been estimated that annually 150 million people were infected with UTI worldwide which costing global economy more than 6 billion US dollars¹. UTI can affect lower and sometimes both lower and upper urinary tracts. The treatment of UTI can be classified into uncomplicated and complicated on the basis of their choice of treatment². UTI is more common in females than in males as female urethra structurally found less effective for preventing the bacterial entry³. It may be due to the proximity of the genital tract

and urethra⁴ and adherence of urothelial mucosa to the mucopolysaccharide lining⁵. The other main factors which make females more prone to UTI are pregnancy and sexual activity⁶. In pregnancy, the physiological increase in plasma volume and decrease in urine concentration develop glycosuria in up to 70% women which ultimately leads to bacterial growth in urine⁷.

The spectrum of bacteria causing complicated UTI is much broader than of those causing uncomplicated UTI. However, the most commonly encountered bacteria are gram negative of which *Escherichia coli*, *Citrobacter* spp., *Enterobacter* spp., *Klebsiella* spp., *Pseudomonas aeruginosa*, and *Proteus* spp., whereas gram

positive *Staphylococcus aureus*, and *Candida* spp. are found rarely¹⁰.

Increasing multidrug resistance in bacterial uropathogens is an important and emerging public health problem. Increasing drug resistance in UTI needs regular monitoring of the antibiotic susceptibility of uropathogens in a particular area. Various factors such as the type of UTI (complicated or uncomplicated), gender, age, and previous history of antibiotic therapy of each UTI patient should also be considered to find out the correct global data on susceptibility¹¹. The distribution of bacteria and their antimicrobial susceptibility data of UTI-causing microorganisms changes from time to time and from place to place⁹. The susceptibility data provided by regional microbiology laboratories helps to choose the empirical choice of antimicrobials to treat UTI; however, these conditions are limited to complicated UTI as the samples of uncomplicated UTI are rarely sent to laboratories^{12,13}. Generally, the antimicrobial treatment is initiated before the laboratory results which may lead to the frequent misuse of antibiotics¹⁴. The resistance pattern of community acquired uropathogens has not been extensively increased in our country¹⁵⁻¹⁷. To the best of our knowledge, no data regarding the bacterial resistance in UTIs from Rangpur city has been documented. Since most UTIs are treated empirically. Therefore there is a need for periodic monitoring of etiologic agents of UTI and their resistance pattern in this community.

This study was undertaken in view of paucity of reports of UTIs in patients of Rangpur city. The aim of the study is to determine the UTI-causing microorganisms distribution and antimicrobial susceptibility will also be determined against most common bacteria.

Materials and Methods

This cross sectional study was carried out in the microbiology laboratory of the Department of

Microbiology Prime Medical College Hospital, Rangpur from January 2013 to December 2013. A total of 494 urine samples from OPD and IPD irrespective of age and sex at Prime Medical College hospital were examined by standard laboratory methods. Relevant history, laboratory record and findings was recorded in a predesigned data sheet and subsequently analyzed manually.

Sample Collection

Clean catch midstream urine samples of 494 patients were collected from each patient into a 20 mL calibrated sterile screw-capped universal container which was distributed to the patients, comprised of 192 males and 302 females, who attended the outpatient departments (OPDs) and IPD of this hospital with clinical evidence of urinary tract infection, determined by treating physicians. The specimens were labeled, transported to the laboratory, and analyzed within 6 hours. All patients were well instructed on how to collect sample aseptically prior to sample collection to avoid contaminations from urethra. Verbal informed consent was obtained from all patients prior to specimen collection. The study was conducted after due ethical approval which was subjected to the hospital administrations.

Sample Processing

A calibrated loop method was used for the isolation of bacterial pathogens from urinary samples. A sterile 4.0 mm platinum wired calibrated loop was used which delivered 0.001 mL of urine. A loopful urine sample was plated on Hi-chrome, Cystine-Lactose-Electrolyte Deficient (CLED) agar, MacConkey agar, and blood agar medium (Hi Media Laboratories, Mumbai, India). The inoculated plates were incubated at 37°C for 24 h and for 48 h in negative cases. The number of isolated bacterial colonies was multiplied by 1000 for the estimation of bacterial load/mL of the urine sample. A specimen was considered positive for UTI if an organism was cultured at a concentration of $\geq 10^5$ cfu/mL or when an organism was cultured

at a concentration of 10^4 cfu/mL and >5 pus cells per high-power field were observed on microscopic examination of the urine¹⁸.

Identification Bacterial Isolates

Identification of both gram positive and gram negative bacterial isolates was done on the basis of their cultural and biochemical characteristics and gram staining smear made from culture colonies. Gram negative bacteria were identified by the standard biochemical tests^{10,19} and Gram positive microorganisms were identified with the corresponding laboratory tests: catalase, coagulase, and subculture on mannitol salt agar for *Staphylococcus aureus*²⁰. Identified and pure isolates were maintained in nutrient agar slants and incubated at 37°C for 24 hrs. The isolates were subcultured periodically.

Antibiotic Susceptibility Testing

Isolates were tested for antimicrobial susceptibility testing by the standard Kirby Bauer's disc diffusion method²¹. Standard inoculums adjusted to 0.5 McFarland was swabbed on Mueller Hinton agar and was allowed to soak for 2 to 5 minutes. After that antibiotic disks were placed on the surface of media and pressed gently. Mueller Hinton agar plates were then incubated at 37°C for 24 h. After 24 h the zone of growth inhibition were measured and interpreted by the recommendations of clinical and laboratory standards²². The following standard antibiotic discs were used for the isolates, amoxicillin, cephadrine, co-trimoxazole, ciprofloxacin, nitrofurantoin, nalidixic acid, ceftriaxone, amoxicillin-clavulonic, ceftazidime, cefuroxime, amikacin, azithromycin, imipenem and vancomycin. Standard strains of *Esch. coli* (ATCC 25922) was used routinely in this study as control.

Results

In total 494 urine samples, 212 samples showed the significant bacterial growth (table-1) which were comprised of 42.9% samples.

Table 1: Result of culture in suspected urine samples from UTI patients.

Number of samples	Culture positive	Culture negative
494	212 (42.9%)	282 (57.1%)

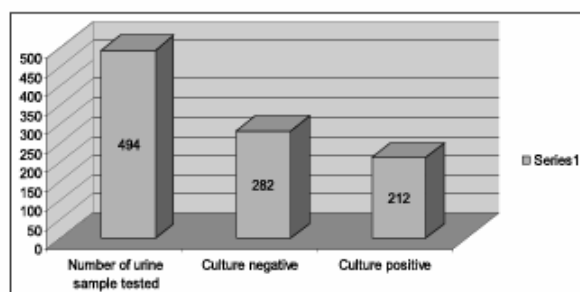


Figure-I: Bacterial isolation rate in the study sample

Among the total of 212 bacterial uropathogens comprised of 200 (94.3%) Gram negative and 12 (5.7%) Gram positive (table-2) were isolated from positive urine samples.

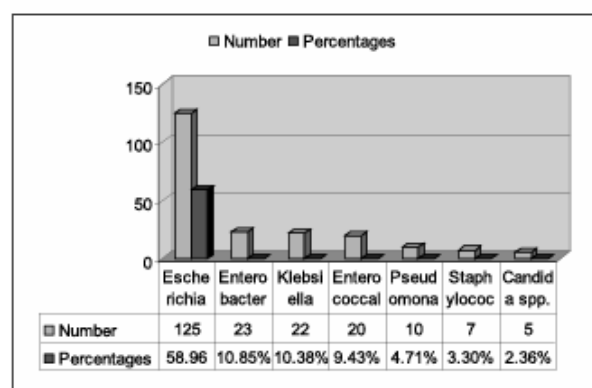
Table 2: Frequency of gram reaction among isolates.

Number of positive samples	Gram negative organisms	Gram positive organisms
212	200 (94.3%)	12 (5.7%)

Escherichia coli (58.96%) was found the dominant bacteria among all isolated uropathogens. The second most prevalent isolate was *Enterobacter* spp. (10.85%), followed by *Klebsiella* spp.(10.38%), *Enterococcal* spp. (9.43%), *Pseudomonas* spp.(4.71%), *Staphylococcus aureus* (3.30%), and *Candida* spp.(2.36%) (table-3).

Table 3: Bacterial distribution in positive culture in all age groups and genders.

Name of Isolates	Number	Percentages
Escherichia coli	125	58.96
Enterobacter spp.	23	10.85
Klebsiella spp.	22	10.38
Enterococcal spp.	20	9.43
Pseudomonas aeruginosa,	10	4.71
Staphylococcus aureus	07	3.30
Candida spp.	05	2.36

**Figure 2:** Bacterial distribution in positive culture in all age groups and genders.

Antibiotic susceptibility results (table-4) showed the resistant and susceptible antibiotics for the tested uropathogen. Overall Ciprofloxacin was found the most resistant drug as 94 (88.4%) against *Escherichia coli*. The second most resistant drug was Nalidixic acid 100 (83.4%) followed by Cefuroxime 94 (78.4%) and Ceftriaxone 93(77.5%); however, the most sensitive drug against *Escherichia coli* was Imipenem (IMP) 113(94.1%) followed by Nitrofurantoin 105(87.5%) and Amikacin 95 (79.0%) (Figure 3).

Table 4: Resistant and susceptibility rates (%) for isolated *Escherichia coli*.

Antibiotic	Sensitive	Resistant
Amoxicillin	32(26.6%)	88(26.6%)
Cephadrine	27(22.8%)	91(77.2%)
Co-trimoxazole	45(37.5%)	75(62.5%)
Ciprofloxacin	26(21.6%)	94(88.4%)
Nitrofurantoin	105(87.5%)	15(12.5%)
Nalidixic acid	20(16.6%)	100(83.4%)
Ceftriaxone	27(22.5%)	93(77.5%)
Amoxicillin-clavulonic acid	77(64.1%)	43(35.9%)
Ceftazidime	38(31.6%)	82(68.4%)
Cefuroxime	26(21.6%)	94(78.4%)
Amikacin	95(79.0%)	25(21.0%)
Azithromycin	57(47.5%)	63(52.5%)
Imipenem	113(94.1%)	07(5.9%)
Vancomycin	36(30.0%)	84(70.0%)

Discussion

This study provides valuable data to compare and monitor the status of antimicrobial resistance against most common uropathogens to improve efficient empirical treatment. Increasing antimicrobial resistance has been documented globally²³⁻²⁶. The prevalence of UTI was found 42.9% in this study and this rate of prevalence is higher than in savar, Dhaka and the other studies in which accounts for 25.6%²⁸, 22%²⁹, 38.6%³⁰, 35.5%⁴¹, 4.2%³¹, 17.19%¹⁶, 10.86%¹⁷, 34.5%³², and 36.68%³³, however, the prevalence rate of UTI in our study correlates with other studies done in South Trinidad³⁴, and in the Mexican population³⁵ which showed such more highly significant uropathogens 49% and 97.3%, respectively.

In this study, the Gram negative bacilli constituted 94.3% of the total bacterial isolates while Gram positive organisms constituted 5.7%. *Escherichia coli* (42.58%) was found the most prevalent gram negative bacteria in the positive urine samples of UTI. This result is consistent with reports from other studies^{32,36,37} but differs from the reports in which *P. aeruginosa*³⁹ and *Klebsiella* spp.⁴⁰ were recorded as the predominant bacteria in UTI. Other isolated bacteria from UTI cases in this study were *Enterobacter* spp. (7.10%), *Klebsiella* spp. (18.71%), *P. aeruginosa* (12.90%), *Staphylococcus aureus* (9.68%), and *Candida* spp.. These findings were not correlate with other reports in which *P. aeruginosa* was reported as the second most common bacterial isolate in UTI studies in India¹⁴ and Lafia, Nigeria⁸; however, these results correlates with others in which *Klebsiella* spp. was reported as the second most frequently isolated organism in UTI^{27,38}. The studies on UTI in other places of the world also showed that *E. coli* and *Klebsiella* spp. are the commonest uropathogens in UTI^{16,17}. Higher incidence of gram negative bacteria, related to *Enterobacteriaceae*, in causing UTI.

Conclusion

Against the background of paucity of reports of UTI in PMC Rangpur, this is the first study conducted to determine the prevalence of UTI, the effect of gender and age on its prevalence, and their susceptibility profile in the community of Rangpur city. This study provides valuable laboratory data to monitor the status of antimicrobial resistance among uropathogens and to improve treatment recommendations in this geographical area.

Acknowledgments

The authors are grateful to the OPD physicians and technical staffs of hospital laboratories for their guidance and efforts. They are also thankful to the hospital administrations for granting them

permission for sample collection from UTI patients and to the Head, Department of Microbiology, PMC, Rangpur, for providing the necessary facilities to carry out this work.

References

1. Gonzalez C M and Schaeffer A J. "Treatment of urinary tract infection: what's old, what's new, and what works". *World Journal of Urology*. 1999;vol 7(6): pp. 372-382.
2. Sabra SM and Abdel MM. "Epidemiological and microbiological profile of nosocomial infection in Taif hospitals, KSA (2010-2011)". *World Journal of Medical Sciences*. 2012;vol7(1): p. 1-9.
3. Warren JW, Abrutyn E, Richard HJ, Johnson JR, Schaeffer AJ, and Stamm WE. "Guidelines for antimicrobial treatment of uncomplicated acute bacterial cystitis and acute pyelonephritis in women". *Journal of Clinical Infectious Diseases*. 1999;vol. 29(4):p. 745-758.
4. Schaeffer AJ, Rajan N, Cao Q et al. "Host pathogenesis in urinary tract infections". *International Journal of Antimicrobial Agents*. 2001;vol. 17(4):p. 245-251.
5. Akortha EE and Ibadin OK. "Incidence and antibiotic susceptibility pattern of *Staphylococcus aureus* amongst patients with urinary tract infection (UTI) in UBTH Benin City, Nigeria". *African Journal of Biotechnology*. 2008;vol. 7(11): p. 1637-1640.
6. Arul KC, Prakasam KG, Kumar D and Vijayan M. "A cross sectional study on distribution of urinary tract infection and their antibiotic utilization pattern in Kerala". *International Journal of Research in Pharmaceutical and Biomedical Sciences*. 2012;vol. 3(3):p. 1125-1130.
7. Lucas MJ and Cunningham FG. "Urinary infection in pregnancy". *Journal of Clinical*

- Obstetrics and Gynecology. 1993;vol. 36(4):p. 855-868.
8. Kolawole AS, Kolawole OM, Kandaki-Olukemi YT, Babatunde SK, Durowade KA, and Kolawole CF. "Prevalence of urinary tract infections (UTI) among patients attending Dalhatu Araf Specialist Hospital, Lafia, Nasarawa State, Nigeria". International Journal of Medicinal Medical Sciences. 2009;vol. 1(5):p. 163-167.
 9. Okonko IO, Ijandipe LA, Ilusanya OA et al. "Incidence of urinary tract infection (UTI) among pregnant women in Ibadan, South-Western Nigeria". African Journal of Biotechnology. 2009;vol. 8(23):p. 6649-6657.
 10. Foxman B and Brown P. "Epidemiology of urinary tract infections: transmission and risk factors, incidence, and costs". Journal of Infectious Disease Clinics of North America. 2003;vol. 17(2):p. 227-241.
 11. Alos JI. "Epidemiology and etiology of urinary tract infections in the community. Antimicrobial susceptibility of the main pathogens and clinical significance of resistance". Enfermedades Infecciosas y Microbiología Clínica. 2005; vol. 23(4):p. 3-8.
 12. McNulty CAM, Richards J, Livermore DM et al. "Clinical relevance of laboratory-reported antibiotic resistance in acute uncomplicated urinary tract infection in primary care". Journal of Antimicrobial Chemotherapy. 2006;vol. 58(5):p. 1000-1008.
 13. Car J. "Urinary tract infections in women: diagnosis and management in primary care". British Medical Journal. 2006;vol. 332(7):p. 94-97.
 14. Tambekar DH, Dhanorkar DV, Gulhane SR, Khandelwal VK, and Dudhane MN. "Antibacterial susceptibility of some urinary tract pathogens to commonly used antibiotics". African Journal of Biotechnology. 2006;vol. 5(17):p. 1562-1565.
 15. Biswas D, Gupta P, Prasad R, Singh V, Arya M, and Kumar A. "Choice of antibiotic for empirical therapy of acute cystitis in a setting of high antimicrobial resistance". Indian Journal of Medical Sciences. 2006;vol. 60(2):p. 53-58.
 16. Akram M, Shahid M, and Khan AU. "Etiology and antibiotic resistance patterns of community-acquired urinary tract infections in J N M C Hospital Aligarh, India". Annals of Clinical Microbiology and Antimicrobials. 2007; vol. 6(4):p. 25-28
 17. Kothari A and Sagar V. "Antibiotic resistance in pathogens causing community-acquired urinary tract infections in India: a multicenter study". The Journal of Infection in Developing Countries. (2008);vol. 2(5):p. 354-358
 18. Collee JG, Miles RS, and Watt B. "Tests for the identification of bacteria: Mackie and Mc Artney Practical Medical Microbiology. ed. 17. London, UK: Churchill Livingstone ; 1996. p. 433-6.
 19. Foxman B, Barlow R, D'Arcy H, Gillespie B, and Sobel JD. "Urinary tract infection: self-reported incidence and associated costs". Annals of Epidemiology. 2000;vol. 10(8):p. 509-515.
 20. Andreua A, Alos JI, Gobernado M, Marco F, Rosa DLM, and Garcia-Rodriguez JA. "Etiology and antimicrobial susceptibility among uropathogens causing community-acquired lower urinary tract infections: a nationwide surveillance study". Enfermedades Infecciosas y Microbiología Clínica. 2005;vol. 23(1):p. 4-9.
 21. Bauer AW, Kirby WM, Sherris JC, and Turck M. "Antibiotic susceptibility testing by a standardized single disk method". American Journal of Clinical Pathology. 1966;vol. 45(4):p. 493-496.

22. Clinical and Laboratory Standards Institute (CLSI) "Document M100-S17. Performance standards for antimicrobial susceptibility testing(2007)," CLSI, 940 West Valley Road, Suite 1400, Wayne, Pennsylvania 19087-1898, USA
23. Jani M, Shah S, and Prajapati S. "Antibacterial screening and qualitative phytochemical estimation of selected aquatic plants". *Advances in Biological Research*. 2012;vol. 6(1):p. 19-23.
24. Kashef N, Djavid GE, and Shahbazi S. "Antimicrobial susceptibility patterns of community-acquired uropathogens in Tehran, Iran". *The Journal of Infection in Developing Countries*.2010;vol. 4(4):p. 202-206.
25. Karlowsky JA, Jones ME, Thornsberry C, Critchley I, Kelly LJ, and Sahm DF. "Prevalence of antimicrobial resistance among urinary tract pathogens isolated from female outpatients across the US in 1999". *International Journal of Antimicrobial Agents*. 2001;vol. 18(2):p. 121-127.
26. Rajalakshmi V and Amsaveni V. "Antibiotic susceptibility of bacterial pathogens isolated from diabetic patients". *International Journal of Microbiological Research*. 2012;vol. 3(1):p. 30-32.
27. Haghi-Ashtiani M, Sadeghifard N, Abedini M, Soroush S, and Taheri-Kalani M. "Etiology and antibacterial resistance of bacterial urinary tract infections in children's medical center, Tehran, Iran". *Acta Medica Iranica*. 2007; vol. 45(2):p. 153-157.
28. Nedolisa. *Bacteriology of Urinary Tract Infection amongst Patients Attending Jos University Teaching Hospital (JUTH) [M.S. thesis] (1998), University of Jos, Jos, Nigeria.*
29. Ekweozor CC and Onyemenen TN. "Urinary Tract Infection in Ibadan, Nigeria: causative organism and anti-microbial sensitivity pattern". *African Journal of Medical Sciences*.1996; vol. 25:p. 165-169.
30. Akinyemi KO, Alabi SA, Taiwo MA, and Omonigbehin EA. "Antimicrobial susceptibility pattern and plasmid profiles of pathogenic bacteria isolated from subjects with urinary tract infections in Lagos, Nigeria". *Nigerian Quarterly Journal of Hospital Medicine*. 1997;vol. 1:p. 7-11.
31. Bigwan EI and Elijah D. "Prevalence of Escherichia coli among uropathogens in asymptomatic bacteriuria in a Nigerian Tertiary School in Jos, Nigeria". *International Journal of Biomedical and Advance Research*. 2013;vol. 4(3):p. 198-202.
32. Dash M, Padhi S, Mohanty I, Panda P, and Parida B. "Antimicrobial resistance in pathogens causing urinary tract infections in a rural community of Odisha, India". *Journal of Family and Community Medicine*.2013;vol. 20(1):p. 20-26.
33. Mehta M, Bhardwaj S, and Sharma J. "Screening of urinary isolates for the prevalence and antimicrobial susceptibility of Enterobacteria other than Escherichia coli". *International Journal of Life Science and Pharma Research*. 2013;vol. 3(1):p. 100-104.
34. Orrett FA. "Urinary tract infections in general practice in a rural community in South Trinidad". *Saudi Medical Journal*.2001;vol. 22(6):p. 537-540.
35. García-Morúa A, Hernández-Torres A, Salazar-de-Hoyos JL, Jaime-Dávila R, and Gómez-Guerra LS. "Community acquired urinary tract infection etiology and antibiotic resistance in a Mexican population group". *Revista Mexicana de Urología*.2009;vol. 69:p. 45-48.
36. Dimitrov TS, Udo EE, Awni F, Emara M, and Passadilla R. "Etiology and antibiotic susceptibility patterns of community-acquired urinary tract Infections in a Kuwait Hospital".

- Medical Principles and Practice.2004;vol. 13(6):p. 334-339.
37. Omigie O, Okoror L, Umolu P, and Ikuuh G. "Increasing resistance to quinolones: a four-year prospective study of urinary tract infection pathogens". International Journal of General Medicine.2009; vol. 2:p. 171-175.
38. Gales AC, Jones RN, Gordon KA et al. "Activity and spectrum of 22 antimicrobial agents tested against urinary tract infection pathogens in hospitalized patients in Latin America: report from the second year of the SENTRY Antimicrobial Surveillance Program (1998)". Journal of Antimicrobial Chemotherapy. 2000;vol. 45(3):p. 295-303.
39. Ehinmidu JO, Bolaji RO, and Adegboye EEA."Isolation and antibiotic susceptibility profile of Neisseria gonorrhoeae isolated from urine samples in Zaria, northern Nigeria". Journal of Phytomedicine and Therapeutics. 2003;vol. 8(11):p. 20-24.
40. Aboderin OA, Abdu LR, Odetoyin BW, and Lamikanra A. "Antimicrobial resistance in Escherichia coli strains from urinary tract infections". Journal of the National Medical Association. 2009;vol. 101(12):p. 1268-1273.